

This article was downloaded by:

On: 26 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713597286>

Potent and Selective Ligands for Adenosine Binding Sites

G. Cristalli^a; E. Camaioni^a; E. Di Francesco^a; A. Eleuteri^a; S. Vittori^a; R. Volpini^a

^a Dipartimento di Scienze Chimiche, Università di Camerino, Camerino, Italy

To cite this Article Cristalli, G. , Camaioni, E. , Francesco, E. Di , Eleuteri, A. , Vittori, S. and Volpini, R.(1997) 'Potent and Selective Ligands for Adenosine Binding Sites', *Nucleosides, Nucleotides and Nucleic Acids*, 16: 7, 1379 — 1388

To link to this Article: DOI: 10.1080/07328319708006189

URL: <http://dx.doi.org/10.1080/07328319708006189>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

POTENT AND SELECTIVE LIGANDS FOR ADENOSINE BINDING SITES

G. Cristalli*, E. Camaioni, E. Di Francesco, A. Eleuteri, S. Vittori and R. Volpini
Dipartimento di Scienze Chimiche, Università di Camerino, 62032 Camerino, Italy

Abstract. A number of selective ligands for the different binding sites of adenosine have been synthesized and tested in several pharmacological models. The aim of these synthetic efforts is both to improve the knowledge of structure-activity relationships in the adenosine-related biological systems and to develop drugs from some of these molecules.

Introduction

The purine nucleoside adenosine acts as a neurotransmitter or neuromodulator through the interaction with at least four cell surface receptors recently classified as A₁, A_{2A}, A_{2B}, and A₃.¹ These receptor subtypes have been cloned and characterized as belonging to the superfamily of receptors with seven transmembrane helices that couple to G proteins.²

In addition adenosine is substrate of many enzymes of the purine metabolism (adenosine deaminase, adenosine kinase, and 5'-nucleotidase), and the adenosine structure is also part of other bioactive molecules (ATP, ADP, cAMP, NAD, FAD, S-adenosyl methionine etc.).³

The clinical usefulness of adenosine has been recognised in the United States in the 1980s⁴ and now adenosine is commonly used in the diagnosis and treatment of supraventricular arrhythmias.⁵ However, the fact that adenosine is present throughout the body in all organs is a severe problems in the development of drugs acting specifically at the different adenosine binding sites.

Therefore our synthetic efforts during the last decade were devoted to the acquisition of structure-activity relationships and to the development of potent and selective agonists for adenosine binding sites. Some of successful ligands synthesized in our laboratory are listed in Figure 1.

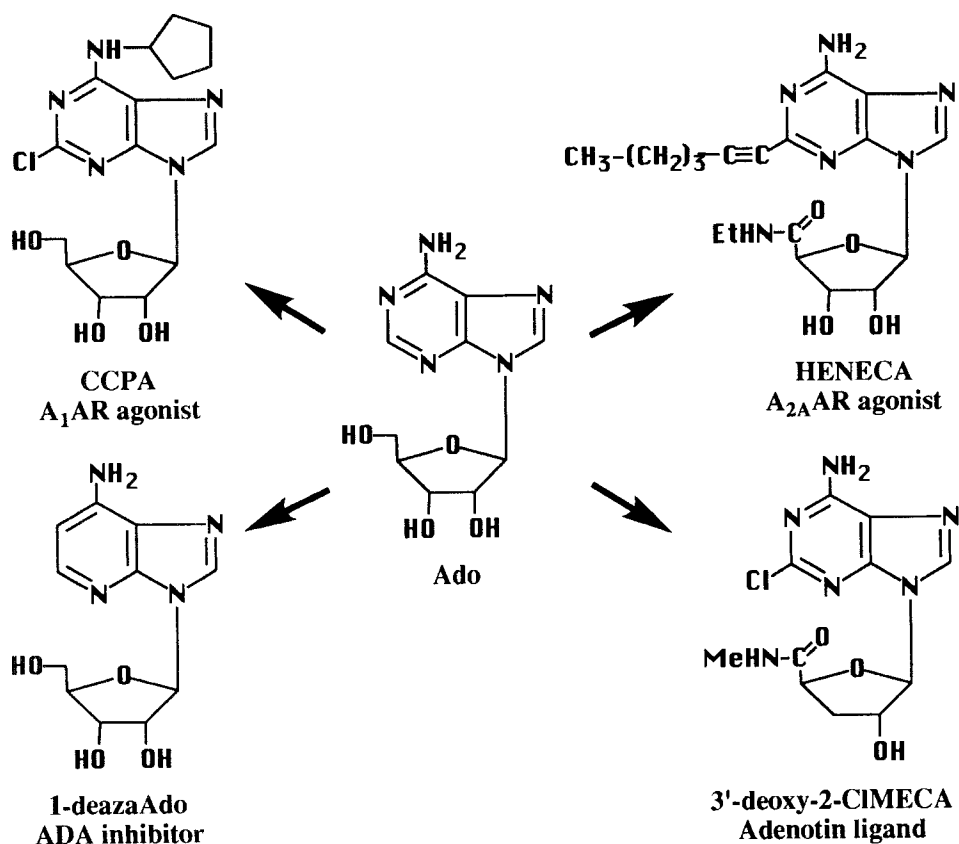


FIGURE 1. Some ligands for adenosine binding sites.

HENECA and *A*_{2A} adenosine receptor agonists

The efforts of medicinal chemists in the past 20 years led to the discovery of a variety of selective agonists for the different adenosine receptor subtypes. A common figure of such agonists is their structural relation with adenosine.

The potent and selective agonists for the *A*_{2A} receptors obtained in the last years were designed on the basis of two observations: a) adenosine-5'-N-ethyluronamide (NECA) has affinity in the low nanomolar range for both *A*_{2A} and *A*₁ receptors,⁶ and is a potent tool in several pharmacological models b) introduction on C-2 of adenosine of a variety of substituents increased *A*_{2A} affinity and selectivity.

On these bases, we have reported the synthesis of the 2-hexynyl derivative of NECA, identified as HENECA, which showed high affinity at *A*_{2A} adenosine receptors and good *A*_{2A} vs *A*₁ selectivity.⁷ HENECA displayed also inhibitory activity on platelet aggregation

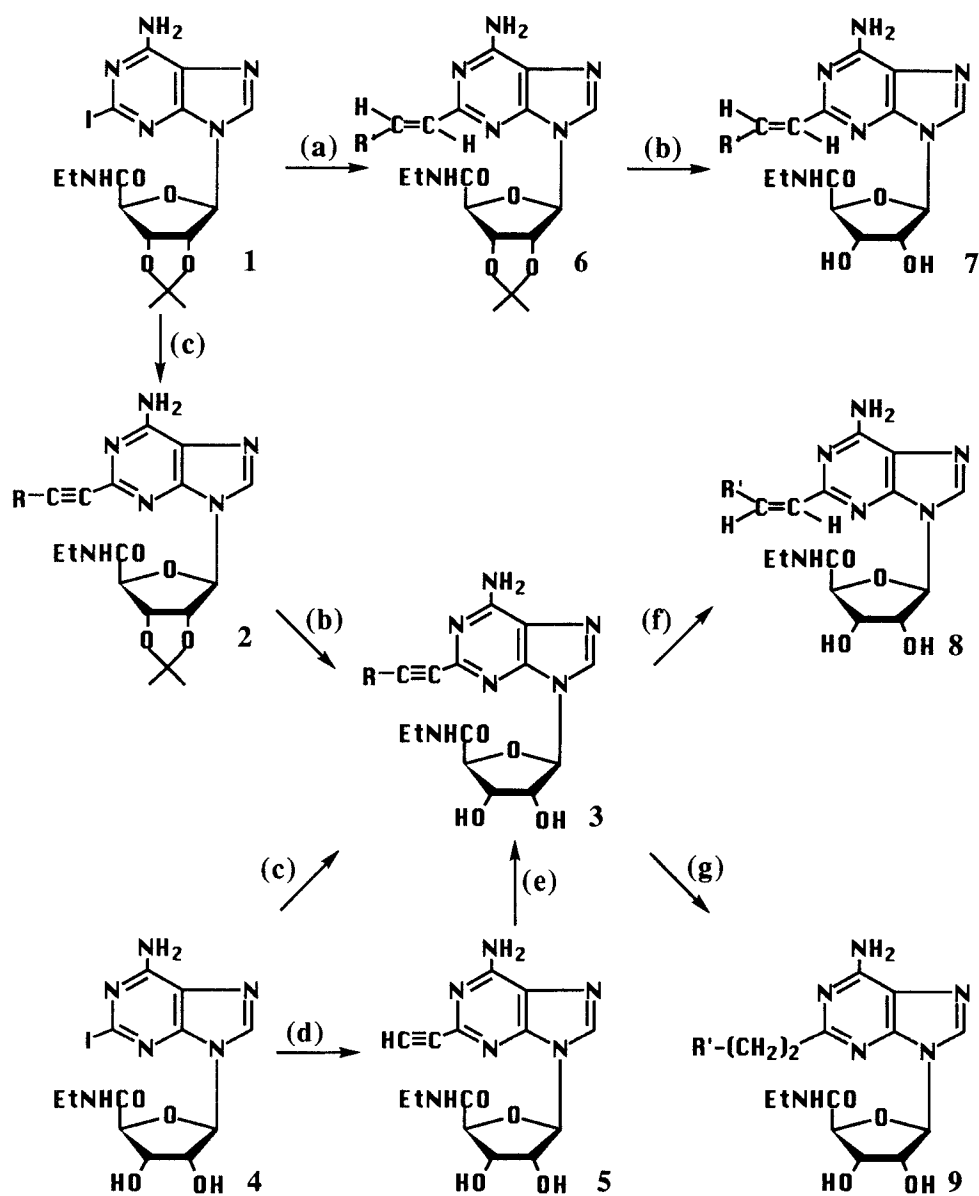
higher than NECA itself,⁸ and has been extensively characterized using a variety of experimental models.^{9,10}

The therapeutic potential of HENECA for the treatment of cardiovascular diseases prompted us to synthesize a number of new 2-alkynyl and cycloalkynyl derivatives of NECA bearing hydroxyl, amino, chloro, cyano and heterocyclic groups or substituted aromatic or heteroaromatic rings in the side chain.^{11,12} Briefly, from the common intermediate, the 5'-carboxyl-2',3'-isopropyliden derivative, the two carboxamido derivatives **1** and **4** were obtained as starting material for the synthetic routes a-d depicted in Scheme 1.¹³ Moreover, some alkenyl and alkyl derivatives of NECA were also synthesized, as classical modification of the triple bond, to better understand the conformational requirements of the receptor area interacting with the substituents in 2- and 5'-position.¹⁴ In the case of the alkenyl derivatives, it is possible to design two geometric isomers: *cis* or *Z* (**8**), and *trans* or *E* (**7**). They were obtained by two different approaches illustrated in Scheme 1. The interaction of the new 2-alkynyl and 2-alkenyl derivatives of NECA with the adenosine receptors was evaluated using both radioligand binding technique and functional assays. In general, many compounds showed subnanomolar activity in the rat striatal binding experiments and some of them were very potent in inducing vasorelaxation without relevant effect on heart rate. In particular, the introduction of a 2-thiazolyl group led to a compound (**11**) with marked vasodilating activity among all the synthesized agonists (rat aorta, $EC_{50} = 3.5$ nM).

Partial reduction of the triple bond in 2-alkynyl derivatives of NECA led to compounds whose activity at A_{2A} receptor subtype was related to *Z-E* isomerism, the *E*-diastereomers being more potent and selective than the *Z* ones. Saturation of the side chain markedly reduced compounds affinity at adenosine receptors. The new nucleoside (*E*)-2-phenylpentenylNECA (**10**) exhibited both high A_{2A} receptor affinity ($K_i = 3.5$ nM) and almost 300-fold A_{2A} vs A_1 selectivity. Comparison between alkenyl derivatives of NECA and adenosine further demonstrated that the 5'-ethylcarboxamido group is critical for the A_{2A} affinity. These studies indicated also that the isomerism of the substituent in 2-position and the nature of the 5'-group in adenosine derivatives are critical to achieve high affinity and selectivity at the A_{2A} adenosine receptor subtype.¹⁴

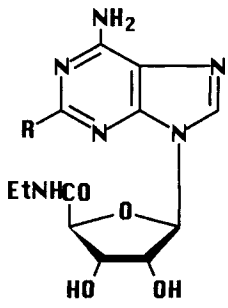
All the synthesized compounds were also tested as inhibitors of platelet aggregation induced by ADP and the results are reported as potency ratio vs NECA.¹¹⁻¹³ Some compounds resulted more potent than HENECA itself as inhibitors of platelet aggregation.

Among these alkynyl derivatives, phenylhydroxypropynylNECA (R,S-PHPNECA) behaved as a very potent agonist at A_1 and A_{2A} receptor subtypes, with a K_i of 2.5 nM and 0.9 nM, respectively. Furthermore, it showed an inhibitory activity on platelet aggregation, induced by ADP, about 16 fold higher than NECA, being the most potent nucleoside inhibitor of platelet aggregation reported so far.

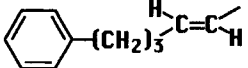


SCHEME 1

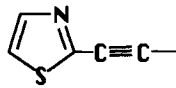
TABLE 1. Pharmacological activity of selected NECA derivatives.



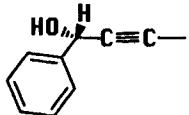
R =



10



11



12

Compds	Binding assays Ki (nM)			Functional activity EC50 (nM)		Antiaggr. act. IC50 (nM)
	Rat brain A ₁	Rat striatum A _{2A}	Selectivity A ₁ /A _{2A}	Rat aorta A ₁	Rat aorta A _{2A}	Rabbit platelets A _{2A}
HENECA	130	2.2	59	>10 μM	596	67
10	1017	3.5	291	>10 μM	576	1400
11	85.4	41.3	2.1	7160	28.7	450
12	4.0	0.5	8	52.0	29.9	6

For experimental details see Ref. 11-13

Since this compound bears a chiral carbon in the side chain, the enantiomeric resolution was undertaken to assess the enantioselectivity of A_{2A} adenosine receptors.¹⁵ The S-diastereomer **12** proved to be more potent than the R-diastereomer, and than the mixture as well, in all model systems. However, the most important achievement is that the platelet receptor clearly discriminates between the two compounds since R-PHPNECA is equiactive with NECA, whereas the S-diastereomer shows an antiaggregatory potency about 40-fold that of NECA, i.e. about 6 nM, resulting the most potent inhibitor so far known in the adenosine agonist derivatives.

CCPA

A potent and selective agonist for the A₁ receptor was designed on the basis that the contemporary presence of a chlorine in 2-position and a cyclopentyl substituent on the N-6 amino group increased both A₁ affinity and selectivity.¹⁶ This observation, related to 1-deazaadenosine derivatives, prompted us to synthesize the 2-chloro derivative of CPA,

namely CCPA.¹⁷ This compound proved to be both more active and A₁-selective than the reference compound, showing A₁ affinity in the subnanomolar range and a selectivity three fold that of CPA. A detailed pharmacological profile of CCPA in binding, functional, and *in vivo* models has been recently reported by Monopoli et al.¹⁸ CCPA is now available in the market also in the tritiated form, obtained by catalytic reduction in a tritium atmosphere of the corresponding cyclopentenyl derivative.¹⁹

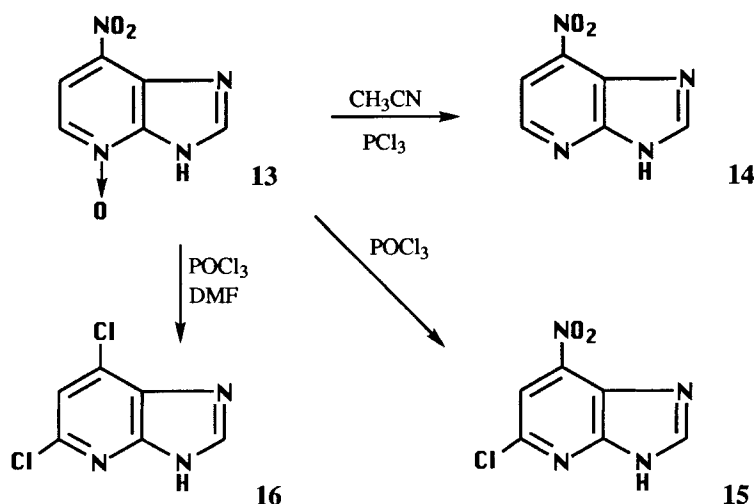
3'-Deoxy-2-CIMECA

Studies performed with the non selective radioligand [³H]NECA have recently demonstrated that in human platelet membranes, in addition to the A_{2A} receptor subtype, a non-receptor protein called "adenotin" is present.²⁰⁻²² Its identity or striking amino-terminal homology with stress-protein sequences and its high intracellular concentration (around 1% of protein in human placenta) suggest that adenotin could play an important role in the regulation of cellular functions like cell proliferation and tumor growth.²³ However, the specific role of adenotin must be determined, yet. Binding studies, performed with typical adenosine ligands, indicated that Ado, N⁶-substituted A₁ adenosine agonists and xanthine antagonists possess very low affinity for this protein.²³ 2-Chloroadenosine-5'-N-methyluronamide (CIMECA) proved to be the most active ligand so far known (K_i = 24 nM). However, it is an unselective ligands, possessing high affinity for both A₁ and A_{2A} adenosine receptors. In an attempt to identify a more potent and selective adenotin ligand we have undertaken the synthesis of 2'- and 3'-deoxy derivative of 2-CIMECA, on the basis that removing of hydroxyl groups from the sugar moiety of adenosine derivatives, while increases binding activity on adenotin, greatly compromises adenosine receptor affinity. Preliminary results of competition experiments for [³H]NECA binding showed that 3'-deoxy-2-CIMECA (Figure 1) possesses very high affinity for adenotin (K_i = 13.7 nM) combined with a very good selectivity toward adenosine receptor subtypes, resulting in the most potent and selective adenotin ligand so far known.²⁴

1-DeazaAdo derivatives

In our effort to synthesize deaza nucleosides endowed with pharmacological activity, we selected 1-deazapurine derivatives since 1-deazaadenosine (1-deazaAdo, Figure 1) has been shown to possess cytotoxic activity,²⁵ to inhibit adenosine deaminase²⁶ and platelet aggregation,¹⁶ and to act as an agonist of adenosine receptors.²⁷

We have already reported the synthesis of 7-nitro-3H-imidazo[4,5-b]pyridine (**14**),²⁵ 5-chloro-7-nitro-3H-imidazo[4,5-b]pyridine (**15**),²⁷ and 5,7-dichloro-3H-imidazo[4,5-b]pyridine (**16**),²⁷ obtained starting from the common intermediate 7-nitro-3H-imidazo[4,5-b]pyridine-4-oxide (Scheme 2).²⁸

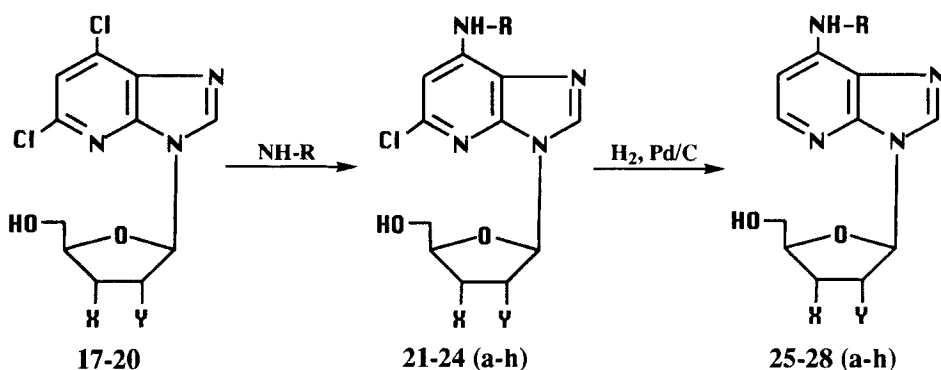


SCHEME 2

The coupling of **2** and **3** with ribose and 2-deoxyribose derivatives gave, after working up, 6-amino-²⁵ and 6-hydroxylamino-1-deazapurine nucleosides.^{29,30} Since 5,7-dichloro-3H-imidazo[4,5-b]pyridine (**16**) is a more versatile base, it has been coupled with a variety of sugar moieties (**17-20**) and transformed in a series of 7-alkyl(aryl)amino derivatives by reacting with the suitable amines (**21-28**, Scheme 3).^{31,32}

These compounds were evaluated for their *in vitro* activity against human immunodeficiency virus type-1 (HIV-1) and herpes simplex virus type-1 (HSV-1).

In addition they were tested for their ability to inhibit adenosine deaminase (ADA) from calf intestine.³² While the parent compounds 1-dezaadenosine (**25a**), 2'-deoxy-1-dezaadenosine (**26a**) and 2',3'-dideoxy-1-dezaadenosine (**27a**) and the corresponding 2-chloro derivatives were inactive, nucleosides bearing cycloalkyl substituents on N⁶ exhibited moderate to good anti-HIV-1 activity, compared to 2',3'-dideoxyadenosine (ddA), with the degree and pattern of improvement depending on the structure of the sugar moiety. In general, 2'-deoxy- and 2',3'-dideoxy derivatives were more potent compounds than the corresponding ribose nucleosides. Compounds bearing a 6-cycloheptyl or cyclooctylamine were the most active in every series. The presence of a chloro group in 2-position improved both activity and therapeutic index in every series, the most active compound being 2'-deoxy-2-chloro-N⁶-cycloheptyl-1-dezaadenosine (**22g**, ED₅₀ = 0.2 μM). On the other hand, most of these derivatives were inactive as anti-HSV-1 agents, showing a high degree of virus-selectivity. The 1-dezaadenine derivatives are not



R = (a) H, (b) CH₃, (c) cC₃H₅, (d) cC₄H₇, (e) cC₅H₉, (f) cC₆H₁₁, (g) cC₇H₁₃, (h) cC₈H₁₅

X = OH, Y = OH; Ribose (17, 21, 25)

X = OH, Y = H; 2'-Deoxyribose (18, 22, 26)

X = H, Y = H; 2',3'-Dideoxyribose (19, 23, 27)

X = H, Y = OH; 3'-Deoxyribose (20, 24, 28)

SCHEME 3

substrates of adenosine deaminase: besides, some of them are good inhibitors of the enzyme. 1-Deaza-2'-deoxyadenosine (**26a**) is the most potent in the series ($K_i = 0.19$ mM) and resulted to be more active than 1-deazaadenosine itself ($K_i = 0.66$ mM). However, the ADA inhibitory activity does not account for the antiviral potency since increased lipophilicity and steric hindrance of substituents resulted in derivatives much less active than the parent compounds. According to the fact that introduction of a chloro group in position 2 of substrates made the compounds more resistant to ADA,³³ the presence of a chloro group in the same position of our compounds produced a decrease in ADA inhibitory activity. This is in agreement with our hypothesis that 1-deazaadenosine derivatives interact with the enzyme directly on the catalytic site, since the presence of a bulky, hydrophobic substituent on the exocyclic nitrogen is detrimental for the hydrogen bonding of the molecules.^{26b}

More recently the synthesis of the 3'-deoxy derivatives of 1-deazaadenosine has been achieved in good yield by coupling compound **16** with 1,2-di-O-acetyl-5-O-benzoyl-3-deoxy-D-ribofuranose³⁴ in the presence of SnCl₄ (**20**).³⁵ The pharmacological activities of the new nucleosides are under investigation.

Acknowledgements

This work was supported by a grant (no 9603341.04) from the Italian Research Council (CNR). We thank M. Brandi, G. Rafaiani and F. Lupidi for technical assistance.

REFERENCES

- 1 a) Fredholm, B.; Abbracchio, M. P.; Burnstock, G.; Daly, J. W.; Harden, T. K.; Jacobson, K. A. Leff, P.; Williams, M. *Pharmacol. Rev.* **1994**, *46* (2), 143-213.
b) Olah, M. E.; Stiles, G. L. *Annu. Rev. Pharmacol. Toxicol.* **1995**, *35*, 581-606.
c) Palmer, T. M.; Stiles, G. L. Adenosine receptors. *Neuropharmacology* **1995**, *34* (7), 683-694.
- 2 Maenhaut, C.; van Sande, S. J.; Libert, F.; Abramowicz, M.; Parmentier, M.; Vanderhaegen, J. J.; Dumont, J. E.; Vassart, G.; Schiffmann, S. *Biochem. Biophys. Res. Comm.* **1990**, *173*, 1169-1178.
- 3 Muller, C. E.; Scior, T. *Pharm. Acta Helv.* **1993**, *68*, 77-111.
- 4 Di Marco, J. P.; Selers, T. D.; Berne, R. M. West, G. A.; Belardinelli, L. *Circulation* **1983**, *68*, 1254-1263.
- 5 Belardinelli, L.; Pelleg, A. In *Purine in cellular signaling: targets for new drugs*. Jacobson, K. A., Daly, J. V., Manganiello, V., Eds.; Springer-Verlag: New York, 1990; pp 95-99.
- 6 Bruns, R. F.; Lu, G. H.; Pugsley, T. A. *Mol. Pharmacol.* **1986**, *29*, 331-346.
- 7 Cristalli, G.; Eleuteri, A.; Vittori, S.; Volpini, R.; Lohse, M. J.; Klotz, K.-N. *J. Med. Chem.* **1992**, *35*, 2363-2368.
- 8 Cristalli, G.; Vittori, S.; Thompson, R. D.; Daly, J. W.; Olsson, R. A. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1994**, *349* (6), 644-650.
- 9 Monopoli, A.; Conti, A.; Zocchi, C.; Casati, C.; Volpini, R.; Cristalli, G.; Ongini, E. *Arzneim. Forsch.* **1994**, *44* (II), 1296-1304.
- 10 Pinna, A.; Wardas, J.; Cristalli, G.; Morelli, M. *Neuroscience*, in press.
- 11 Cristalli, G.; Volpini, R.; Vittori, S.; Camaioni, E.; Monopoli, A.; Conti, A.; Dionisotti, S.; Zocchi, C.; Ongini, E. *J. Med. Chem.* **1994**, *37*, 1720-1726.
- 12 Cristalli, G.; Volpini, R.; Vittori, S.; Camaioni, E.; Monopoli, A.; Conti, A.; Dionisotti, S.; Zocchi, C.; Ongini, E. *J. Med. Chem.* **1995**, *38*, 1462-1472.
- 13 Cristalli, G.; Camaioni, E.; Elsevier Science Publishers, Amsterdam, Pharmacology Chemistry Library, D. Giardinà, S. Piergentili, and M. Pignini Eds, pp 165-180.
- 14 Vittori, S.; Camaioni, E.; Di Francesco, E.; Volpini, R.; Monopoli, A.; Dionisotti, Di Francesco, E.; Volpini, R.; Vittori, S. In *Perspectives in Receptor Research*, 199S.; Ongini, E.; Cristalli, G. *J. Med. Chem.* **1996**, in press.
- 15 Camaioni, E.; Di Francesco, E.; Vittori, S.; Volpini, R.; Monopoli, A.; Casati, C.; Ongini, E.; Cristalli, G. *J. Med. Chem.* submitted.
- 16 Cristalli, G.; Franchetti, P.; Grifantini, M.; Vittori, S.; Klotz, K-N; Lohse, M.J. *J. Med. Chem.* **1988**, *31*, 1179-1183.
- 17 Lohse, M. J.; Klotz, K.-N.; Schwabe, U.; Cristalli, G.; Vittori, S.; Grifantini, M. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1988**, *337*, 687-689.
- 18 Lohse, M. J.; Klotz, K.-N.; Schwabe, U.; Cristalli, G.; Vittori, S.; Grifantini, M. *Naunyn-Schmiedeberg's Arch Pharmacol* **1989**, *340*, 679-683.
- 19 Monopoli, A.; Conti, A.; Dionisotti, S.; Casati, C.; Camaioni, E.; Cristalli, G.; Ongini, E. *Arzneim-Forsch/Drug Res* **1994**, *44* (12), 1350-1312.
- 20 Lohse, M. J.; Elger, B.; Lindenborn-Fotinos, F. J.; Klotz, K.-N.; Schwabe, U. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1988**, *337*, 64-68.
- 21 Hutchison, K. A.; Nevins, B.; Perini, F.; Fox, I. H. *Biochemistry* **1990**, *29*, 5138-5144.
- 22 Lorenzen, A.; Sigrid, G.; Heidrun, V.; Schwabe, U. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1992**, *346*, 63-68.

- 23 Fein, T.; Schulze, E.; Bar, J.; Schwabe, U. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1994**, *349*, 374-380.
- 24 Volpini, R.; Camaioni, E.; Di Francesco, E.; Vittori, S.; Fein, T.; Schwabe, U.; Cristalli, G. *Collect. Czech. Chem. Commun.* **1996**, *61*, S33-S37.
- 25 Cristalli, G.; Franchetti, P.; Grifantini, M.; Vittori, S.; Bordoni, T.; Geroni, C. *J. Med. Chem.* **1987**, *30*, 1686-1688.
- 26 a) Lupidi, G.; Cristalli, G.; Marmocchi, F.; Riva, F.; Grifantini, M. *J. Enzyme Inhib.* **1985**, *1*, 67-75. b) Cristalli, G.; Eleuteri, A.; Vittori, S.; Volpini, R.; Camaioni, E.; Lupidi, G. *Drug Dev. Res.* **1993**, *28* (3), 253-258.
- 27 Cristalli, G.; Grifantini, M.; Vittori, S.; Balduini, W.; Cattabeni, F. *Nucleosides & Nucleotides* **1985**, *4* (5), 625-639.
- 28 Antonini, I.; Cristalli, G.; Franchetti, P.; Grifantini, M.; Martelli, S.; Petrelli, F. *J. Pharm. Sci.* **1984**, *73*, 366-369.
- 29 Cristalli, G.; Vittori, S.; Eleuteri, A.; Grifantini, M.; Volpini, R.; Lupidi, G.; Capolongo, L.; Pesenti, E. *J. Med. Chem.* **1991**, *34*, 2226-2230.
- 30 Cristalli, G.; Vittori, S.; Eleuteri, A.; Grifantini, M.; Lupidi, G.; Capolongo, L.; Pesenti, E. *Nucleosides & Nucleotides* **1991**, *10*(1-3), 253-257.
- 31 Cristalli, G.; Vittori, S.; Eleuteri, A.; Volpini, R.; Camaioni, E.; Lupidi, G. *Nucleosides & Nucleotides* **1994**, *13* (1-3), 835-848.
- 32 Cristalli, G.; Vittori, S.; Eleuteri, A.; Volpini, R.; Camaioni, E.; Lupidi, G.; Mahhmood, N.; Bevilacqua, F.; Palù, G. *J. Med. Chem.* **1995**, *38*, 4019-4025.
- 33 a) Clarke, D. A.; Davoll, J.; Philips, F. S.; Brown, G. B. *J. Pharmacol. Exp. Ther.* **1952**, *106*, 291-302. b) Rockwell, M.; Maguire, M. H. *Mol. Pharmacol.* **1966**, *2*, 574-584.
- 34 Rizzo, C. J.; Dougherty, J. P.; Breslow, R. *Tetrahedron Lett.* **1992**, *33*, 4129-32.
- 35 Volpini, R.; Camaioni, E.; Di Francesco, E.; Vittori, S.; Cristalli, G. submitted.